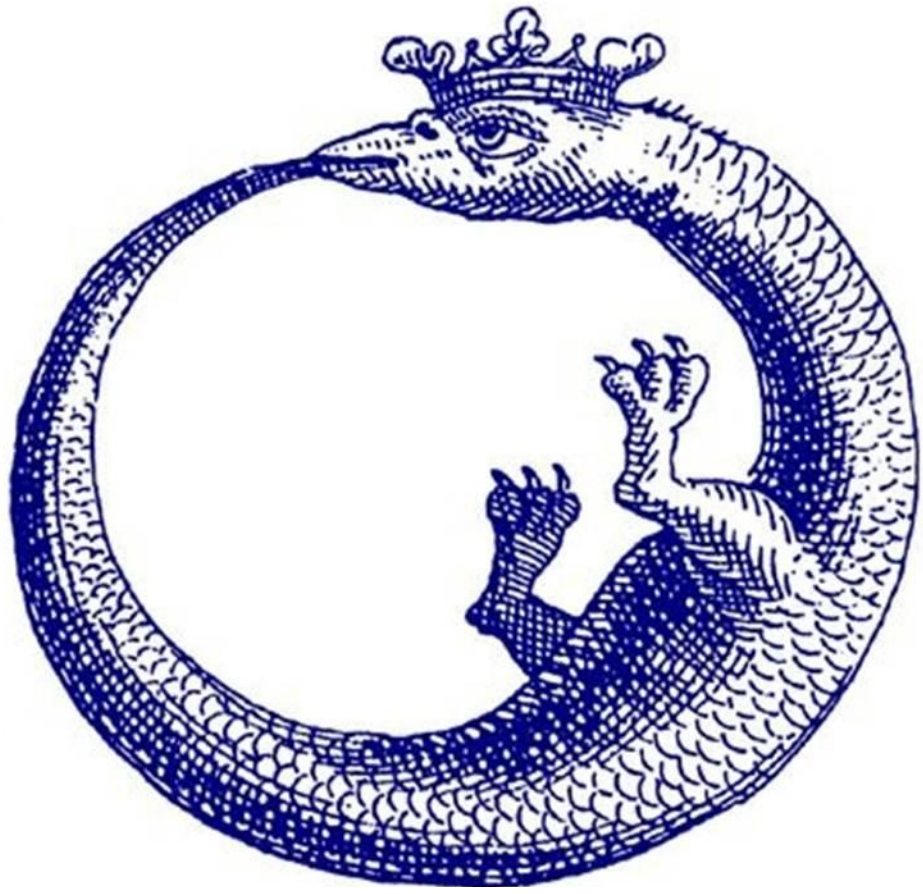


**OROBOROS
S U P P O R T**





- O2k-Workshops
- Videosupport >> *new!*
- wiki.oroboros.at
- **Technical support**



Technical support team



Ondrej
Capek



Gerhard
Krumschnabel

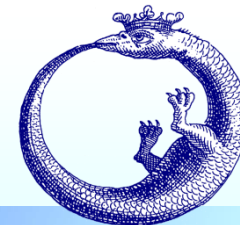


Mona
Fontana-Ayoub

Email us: instruments@oroboros.at

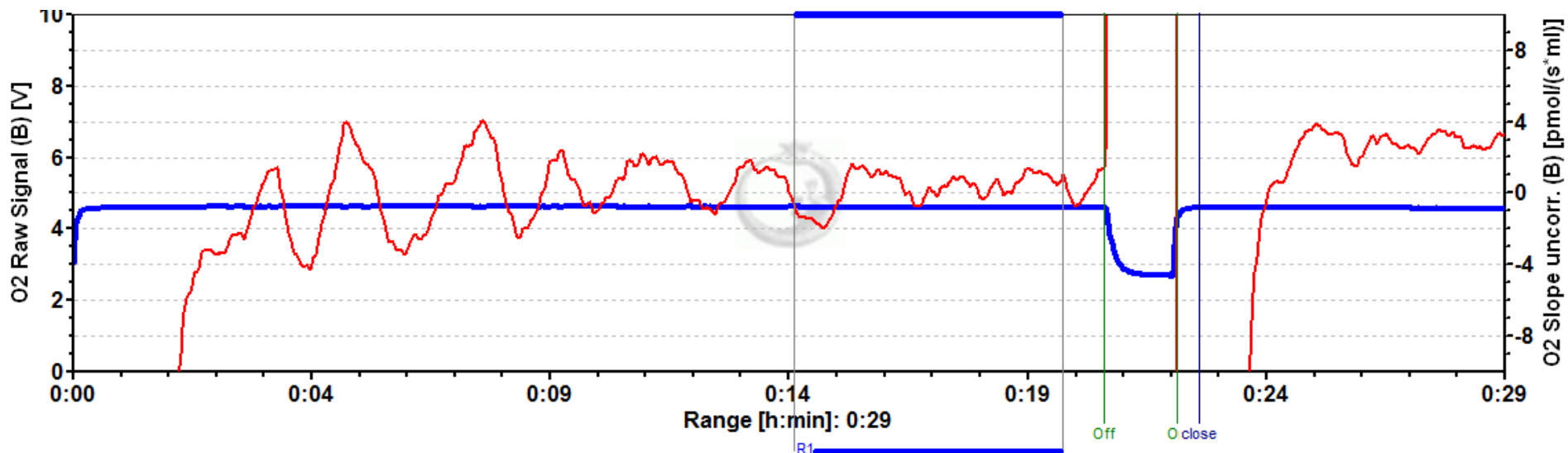


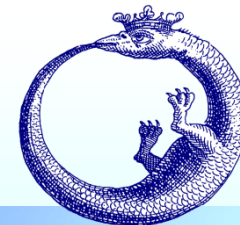
- Avoid problems ;-)
- Try to localize the problem
- Solve them on your own
- Support by OROBOROS



Daily routine before starting an experiment:

- air calibration
- stirrer test
- “medium test”: closed chamber without sample





Regular tests:

- sensor test
- instrumental oxygen background

Sensor test

When?

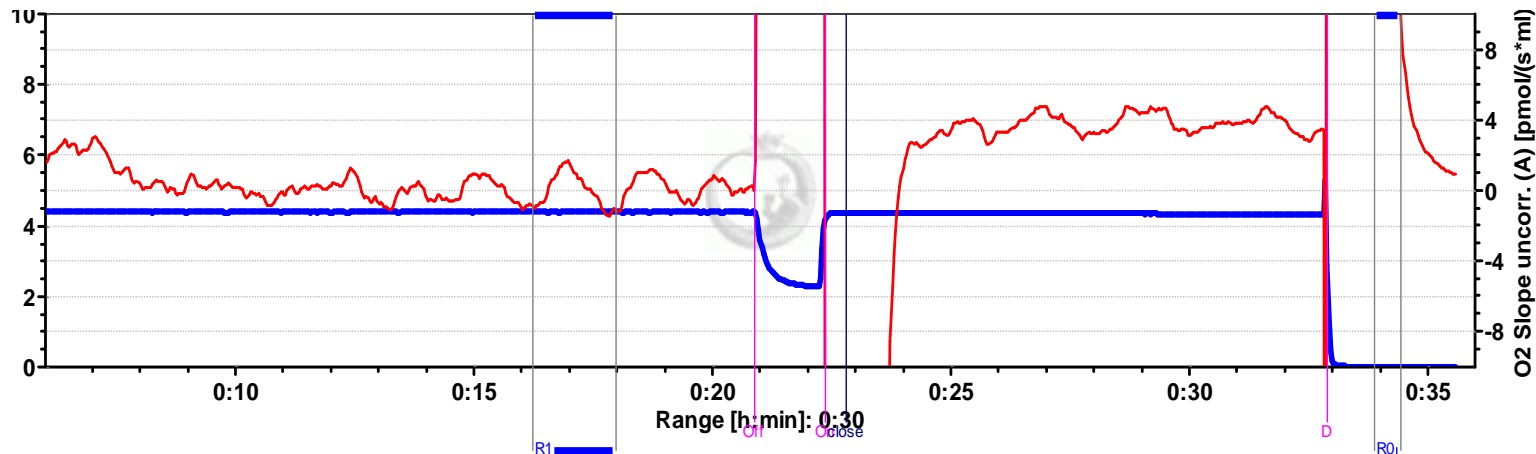
- after sensor service, new membrane,
- routine check
- technical support

Info:

- wiki.oroboros.at „sensor test“
- Demo File: -link from wiki.oroboros.at
 - USB
 - DatLab directory: DLDemo



How to do a sensor test:



Parameters:

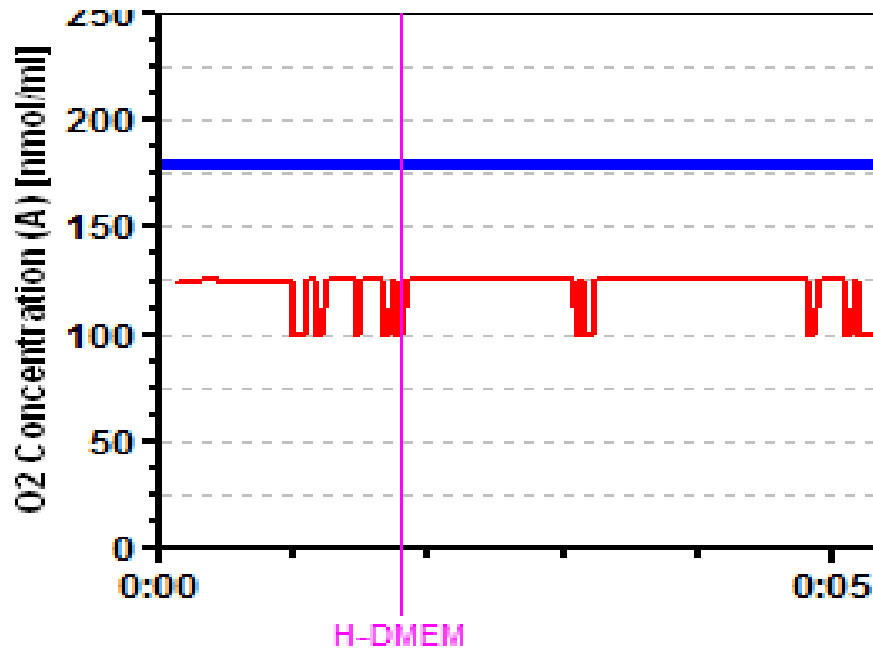
water, $T = 37^{\circ}\text{C}$, gain = 1, stirring = 750 rpm

Procedure:

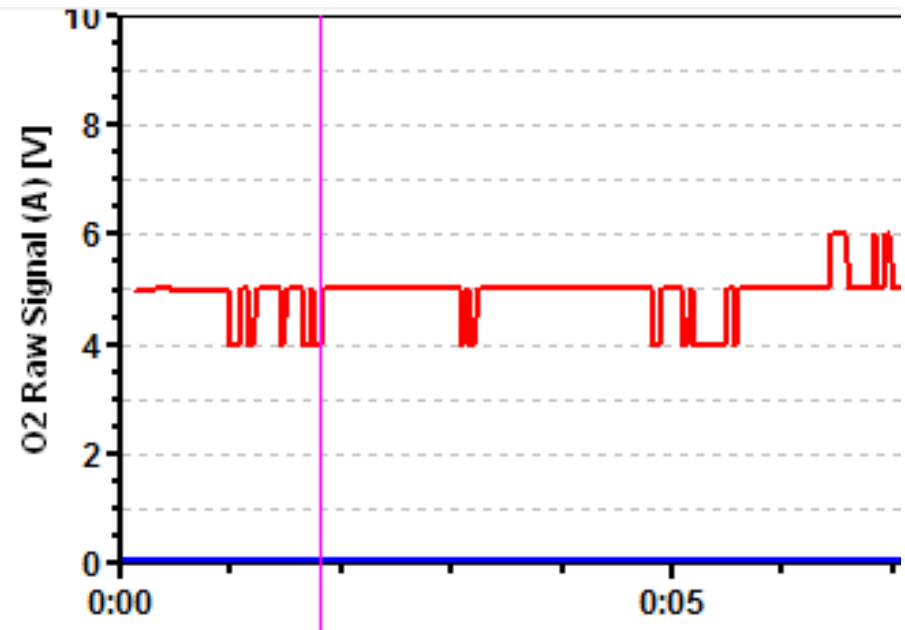
1. use layout “Z Troubleshooting” (raw signal)
2. Air calibration: open chamber, wait for thermal equilibration (stable peltier power)
3. Stirrer test
4. Close chamber (flux up to $\pm 4 \text{ pmol}/(\text{s}\cdot\text{ml})$)
5. Zero calibration (with “Zero solution powder“- dithionite)



Why to look at the raw signal ?



A strange signal

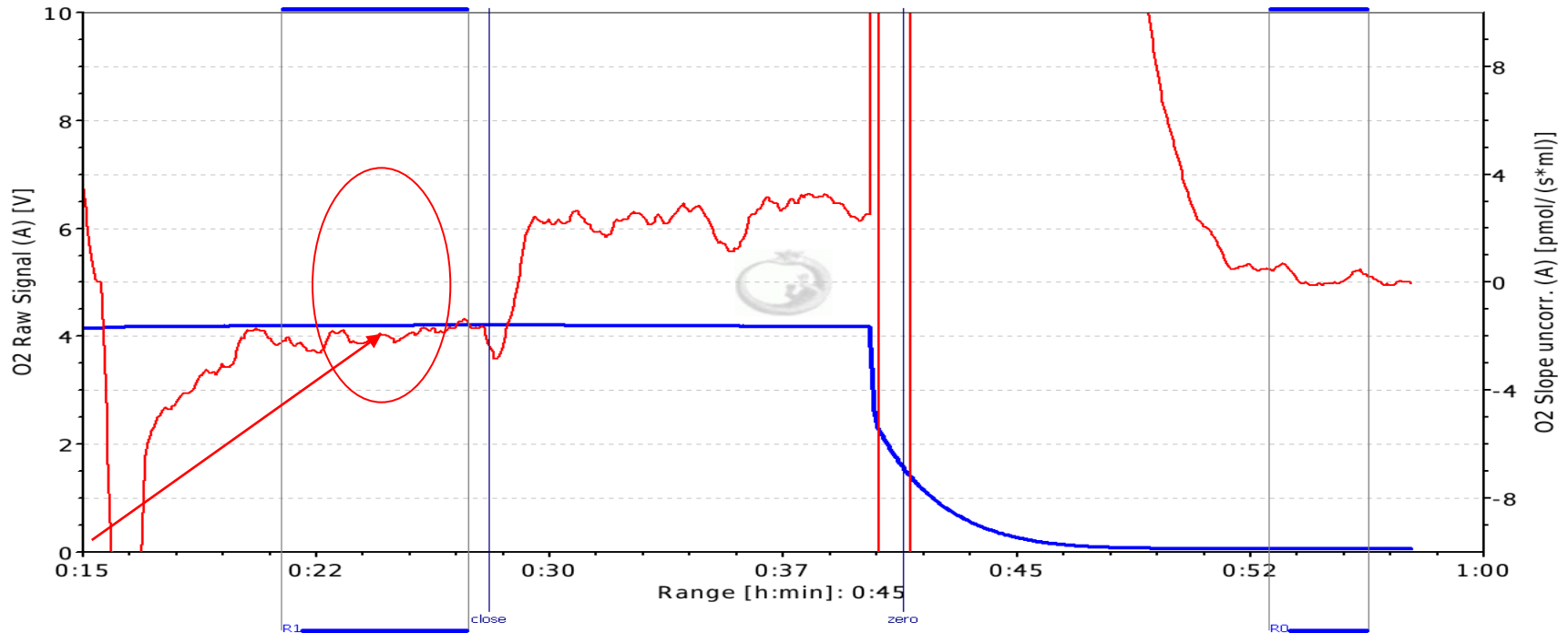


is actually no signal !



O2k technical support - sensor

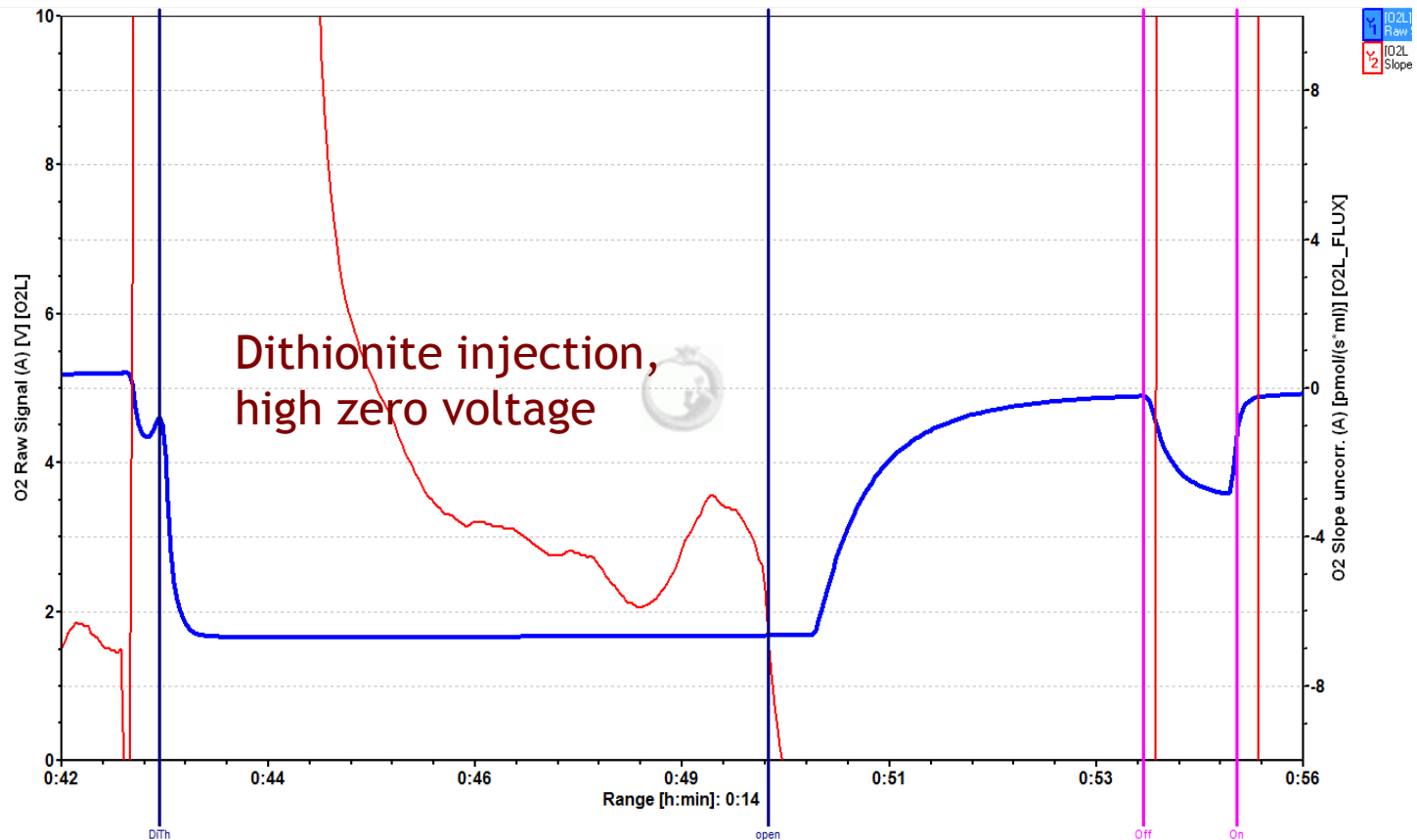
1. Drift of the oxygen signal during calibration - bubbles (sensor, capillary)
2. Slow response

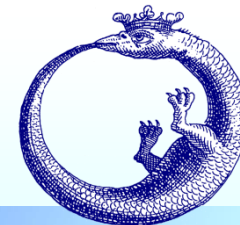




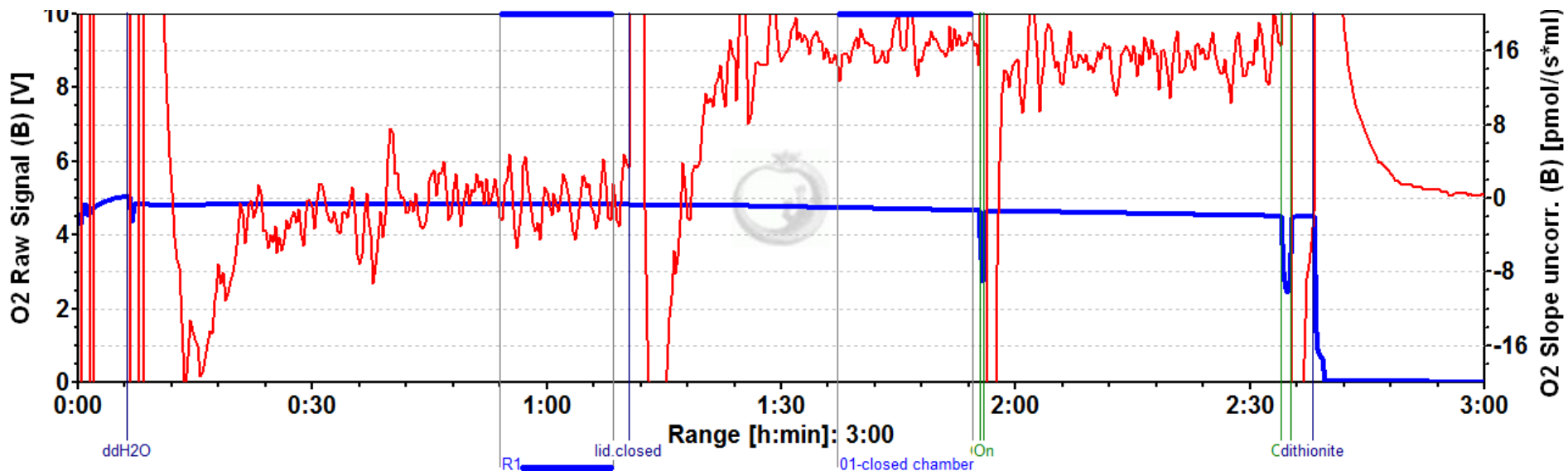
O2k technical support - sensor

Zero current is high (>2.5 %)
Slow response





O2k technical support - biological contamination



normal flux after closing the chamber - up to 4 pmol/(s*ml)

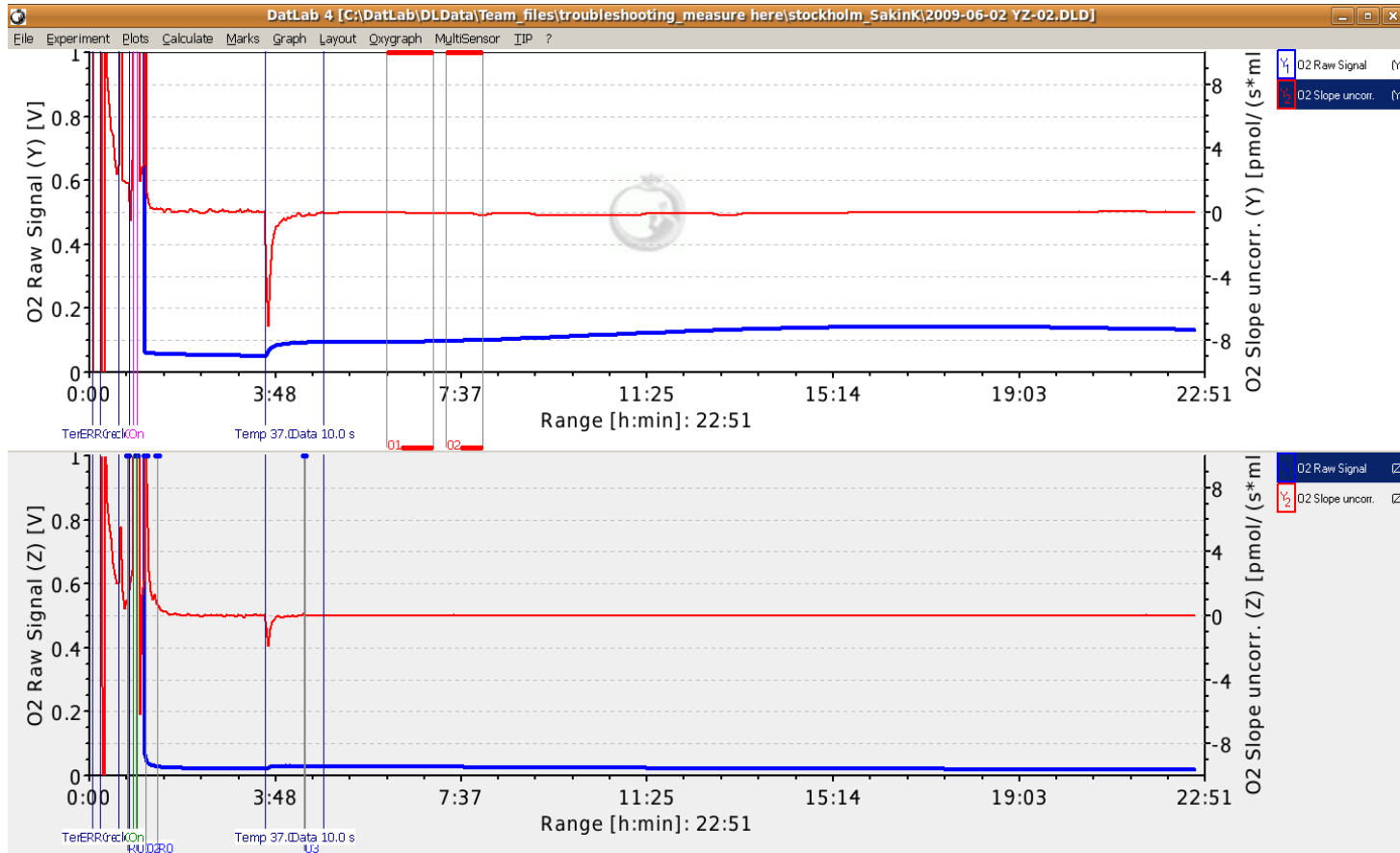
higher values - medium or chamber is contaminated

check if problem persists in water:

NO: new medium

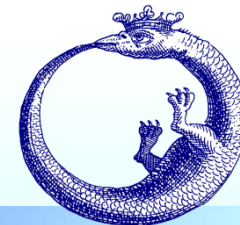
YES: intensiv cleaning of the system with 70 % EtOH

clean glass chamber with 10N HCl



High zero current with drift in the left chamber

Problem was located on sensor connector



O2k technical support - component test

Localization of a Problem

intern.wiki.orooboros

“O2k-technical support”

IN GENERAL

Make a sensor test

Change single components between chambers

sensors

POS holder

glass chamber

stirrers

Run protocol of the sensor test again after switching one component between chambers

If problem occurs now in the other chamber - problem located



Problems with the sensor:

Change membrane

Sensor service:

cathode and anode cleaning

long ammonia service (over night): apply membrane

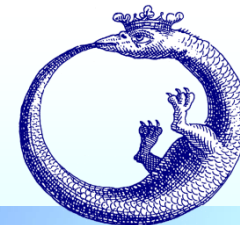
run over night in water before a new test run!

Sensor connector:

Clean the gold pin and threads (water and Methanol/EtOH abs.)

Apply contact oil

MiPNet 08.04 Service of the polarographic oxygen sensor OroboPOS



O2k technical service - FAQ/problems

Stirrer sticks/jumps

- exchange stirrers between chambers
- remove chamber, control for small glass pieces
- clean stirrer and clean chamber and with 10 N HCl

Instrumental Background

- the ultimate instrument test!
- after a new chamber assembly
- before or after a series of experiments (e.g. diagnostics)
- performed in MiR05
- in the oxygen range of your experiment (normoxia, hypoxia...)
- at the experimental temperature



O2k technical service - syringes

TIP - syringes

- TIP2k-Manual: **MiPNet12.10_TIP2k-Manual.pdf**
- Rinse the outside with water immediately after use
- Wash 3x with last used solvent, rinse with EtOH
- Storage: dry
- Rinse with pure solvent before use

Hamilton syringes

- Separate uncoupler and inhibitors from substrates
- Between two runs during the day : rinse outside with water
- End of the day: 3x solvent, 3 x EtOH 100%
- Storage: dry
- **MiPNet19.14 SOP Hamilton microsyringes**
- http://wiki.orooboros.at/index.php/Titration_Set



O2k technical service - chamber cleaning

Chamber cleaning:

- Siphon off the cell/mitochondrial suspension
- Rinse the stoppers and chamber with distilled water five times (fill up to the rim)
- Clean bottom of the stopper and stirrer bar with Kimwipe and rinse with water
- Pptional: wash with remaining cell suspension/isolated mitochondria or tissue to get rid e.g. of sodium azide
- Fill with **70 % EtOH** and insert the stopper making sure that the ethanol fills up the receptacle and cover with perspex cover, leave for 5 min - repeat two more times
- Fill with **EtOH absolute** and leave for 15 min
- Store in 70 % ethanol
- **MiPNet19.03_O2k-cleaning_and_ISS**



O2k technical support - summary

Recommendations:

- Keep your system clean
- During troubleshooting: Discharge yourself, especially before touching the connector - **NO** Crocs
- Try to **localize your problem**

Perform sensor test for:

- your ‘troubleshooting’
- When support by OROBOROS Instruments required
- send us the .DLD file (no screenshot!)

wiki.oroboros.at

Technical Support