

## 117<sup>th</sup> Workshop on high-resolution respirometry & O2k-Fluorometry

2016 September 26-30  
Kuala Lumpur, MY

### Host institution:

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### OROBOROS distributor:

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### Lecturer and tutor:

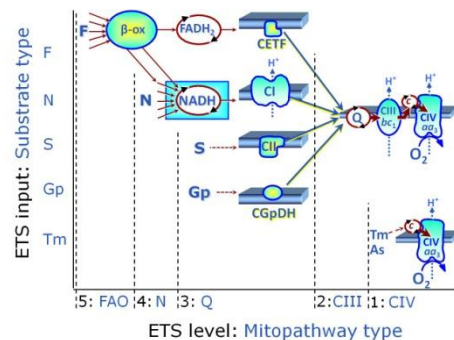
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### Assisted by:

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The **117<sup>th</sup> O2k-Workshop** on high-resolution respirometry and O2k-Fluorometry is held in cooperation with our exclusive distributor in Malaysia. This O2k-Workshop presents a basic introduction to the **OROBOROS Oxygraph-2k** with integrated real-time data analysis. We introduce the new software **DatLab 7** and the concept of a quality control system including the MitoFit interlaboratory proficiency test.

HRR provides information on cell respiration with basic coupling control protocols. State-of-the-art OXPHOS analysis is extended using mt-preparations (permeabilized muscle fibres, tissue homogenate, isolated mitochondria), to evaluate coupling efficiencies and OXPHOS capacities with electron transfer into the Q-junction converging from NADH, FADH<sub>2</sub>, succinate and  $\alpha$ -glycerophosphate (N, F, S, Gp), to diagnose defects in respiratory electron transfer system pathways and the phosphorylation system. Novel developments are presented on **substrate-uncoupler-inhibitor titration (SUIT) protocols** in HRR using the **O2k-Fluorescence LED2-Module** for simultaneous measurement of hydrogen peroxide production (Amplex red<sup>®</sup>). Discussions are extended to measurement of mt-membrane potential using Safranin (fluorometric), and on perspectives of HRR in mitochondrial physiology.



## Programme

<b>1</b>	<b>Monday, Sep 26</b>
<b>Workshop Day 1</b>	
<b>15:00</b>	<b>Local arrangements</b>
<b>15:30-17:30</b>	O2k instrumental setup with the distributor.
<b>2</b>	<b>Tuesday, Sep 27</b>
<b>Workshop Day 2</b>	
<b>09:00-09:45</b>	<b>Welcome</b> , introduction of participants. <b>Videoclips:</b> Overview of O2k setup.
<b>09:45-10:30</b>	Hands-on: Oxygen sensor service and O2k setup.
10:30	<i>Coffee break</i>
<b>10:45-12:00</b>	<b>Instrumental quality control 1:</b> O <sub>2</sub> calibration and the O2k quality control system.
12:00	<i>Lunch break</i>
<b>13:00-15:30</b>	<b>O2k-Fluo experiment:</b> Simultaneous measurement of oxygen consumption (O2k-Core) and H <sub>2</sub> O <sub>2</sub> production (O2k-Fluo LED2-Module) in intact cells (yeast).
15:30	<i>Coffee break</i>
<b>15:45-17:30</b>	<b>DatLab guide</b> through the menus and DatLab O <sub>2</sub> flux analysis: the new DatLab 7 version.
<b>3</b>	<b>Wednesday, Sep 28</b>
<b>Workshop Day 3</b>	
<b>09:00-10:00</b>	<b>O2k-Fluo and hydrogen peroxide production:</b> from isolated mitochondria to tissue homogenate and intact cells.
<b>10:00-10:30</b>	<b>Getting started</b> with an O2k experiment: washing, stirrer test, air calibration.
10:30	<i>Coffee break</i>
<b>10:45-12:00</b>	<b>Experimental set-up:</b> Calibration of O2k-Fluo Sensors.
12:00	<i>Lunch break</i>
<b>13:00-15:30</b>	<b>O2k-experiment:</b> Neuroblastoma – intact cells (ce) vs. permeabilized cells (pce, mt-preparation).
15:30	<i>Coffee break</i>
<b>15:45-17:30</b>	<b>SUIT protocol</b> and DatLab analysis with Excel templates: Flux per volume, flux per mass, flow per cell, flux control ratio, flux control factor.
<b>4</b>	<b>Thursday, Sep 29</b>
<b>Workshop Day 4</b>	
<b>09:00-10:30</b>	<b>O2k-Fluo and mt-membrane potential:</b> calibration, SUIT protocols, problems and solutions.
10:30	<i>Coffee break</i>
<b>10:45-12:00</b>	<b>O2k-technical support and the OROBOROS website.</b> <b>Instrumental quality control 2:</b> O <sub>2</sub> background test with the TIP2k – automatic run over lunch.
12:00	<i>Lunch break</i>
<b>13:00-15:30</b>	O <sub>2</sub> background analysis. <b>Open questions – the modular O2k concept:</b> From O2k-Core applications to advanced O2k-MultiSensor analysis.
15:30	<i>Coffee break</i>
<b>15:45-17:30</b>	<b>Open questions – mitochondrial physiology:</b> experimental design and SUIT protocols.

<b>5</b>	<b>Friday, Sep 30</b>
<b>Workshop Day 5</b>	
<b>09:00-10:30 Open questions – DatLab analysis.</b>	
10:30 <i>Coffee break</i>	
<b>10:45-12:00 Summary – feedback - next steps.</b>	

## List of participants

Participant	Institution
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**Recommended reading**

Gnaiger E (2008) Polarographic oxygen sensors, the oxygraph and high-resolution respirometry to assess mitochondrial function.  
 In: Mitochondrial Dysfunction in Drug-Induced Toxicity (Dykens JA, Will Y, eds) John Wiley:327-52.

»[Full text in Bioblast](#)«



**O2k-Core Manual:**

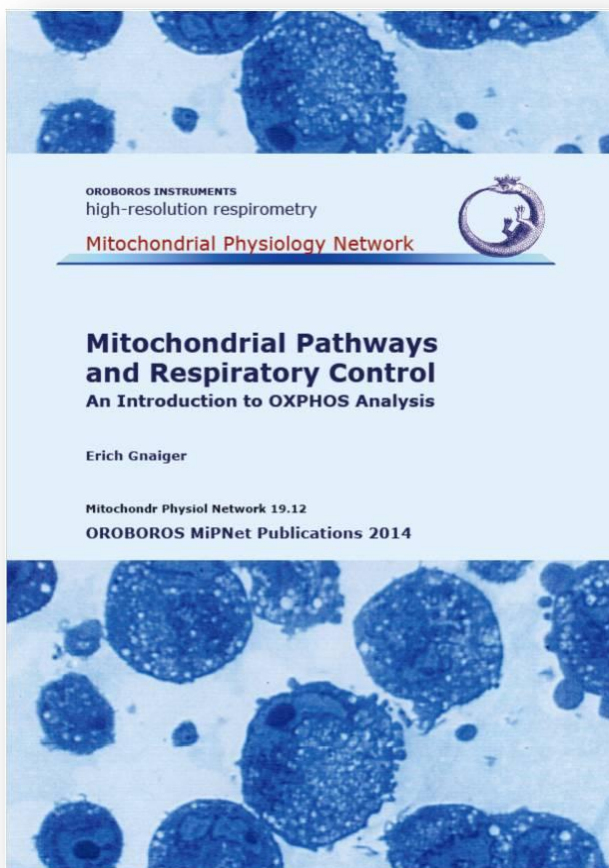
»[Full text in Bioblast](#)«



**SUIT protocols for O2k high-resolution respirometry**

Gnaiger E (2014) Mitochondrial pathways and respiratory control. An introduction to OXPHOS analysis. 4th ed. Mitochondr Physiol Network 19.12. OROBOROS MiPNet Publications, Innsbruck:80 pp.

»[Full text in Bioblast](#)«



**HRR and O2k-Fluorometry**

»[Manual: O2k-Fluo LED2-Module](#)«

Makrecka-Kuka M, Krumschnabel G, Gnaiger E (2015) High-resolution respirometry for simultaneous measurement of oxygen and hydrogen peroxide fluxes in permeabilized cells, tissue homogenate and isolated mitochondria. Biomolecules 5:1319-38. »[Bioblast link](#)«

Krumschnabel G, Eigentler A, Fasching M, Gnaiger E (2014) Use of safranin for the assessment of mitochondrial membrane potential by high-resolution respirometry and fluorometry. Methods Enzymol 542:163-81. »[Bioblast link](#)«

»[O2k-Fluorometry Publications](#)«



**COST Action CA15203 Mitochondrial fitness mapping**

**MITOEAGLE:** Evolution - Age - Gender - Lifestyle - Environment



Contribution to K-Regio project **MitoFit**.

The project MitoFit is funded by the Land Tirol within the program K-Regio of Standortagentur Tirol. [www.mitofit.org](http://www.mitofit.org)



Standortagentur