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Laboratory Protocol: Isolation of Rat Heart Mitochondria

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Preparation: Switch on centrifuge and let it cool down to 4 °C. Keep all buffers, dissection gear, homogenization tools and centrifuge rotor at 4 °C (or on ice).

Anesthesia: Rats are anesthetised by intraperitoneal injection of Thiopental (0.1 g/kg) or by CO₂ narcosis.

Isolation procedure:

- 1. kill rat, dissect out heart (take weight) and put it into ice-cold isolation medium A, wash to remove blood, discard all medium
- 2. cut the heart into small pieces (should become a mash), add drops of isolation buffer A while cutting
- 3. add isolation buffer B (10 ml/g tissue), transfer to pre-cooled glass/Teflon potter and homogenize with 8-12 strokes at medium speed (1000 rpm)
- 4. transfer the homogenate to a 50 ml beaker, bring volume up to ~ 15 ml/g tissue with isolation buffer A, place it on a magnetic stirrer, stir slowly for 20 min in an ice bath
- 5. re-homogenize the homogenate briefly in the potter, bring the volume up to ~ 20–30 ml with isolation buffer C
- 6. centrifuge: 1000 x g, 10 min, 4°C
- 7. transfer the supernatant to a new 50 ml falcon tube
- 8. centrifuge: 6200 x g, 10 min, 4°C
- discard the supernatant, carefully re-suspend the mitochondrial pellet in a small volume of isolation buffer C and bring the volume to 20-30 ml (for 1 g tissue) with isolation buffer C
- 10. centrifuge: 6200 x g, 10 min, 4°C
- 11. discard supernatant and carefully re-suspend mitochondria with small volume of isolation buffer C (the volume of mitochondrial suspension for 1 g tissue ~ 1.2 ml)
- 12. store mitochondria in ice/water bath, use within 3-4 h
- 13. transfer subsamples (20 μ l) into Eppendorf tubes and store at 20°C for further analysis (protein concentration, citrate synthase)

14. for respiration measurements add 2.5 μl of mitochondrial suspension into a 2 ml chamber

Isolation buffer A:

Chemical	Final conc.	Required for
		500 ml buffer
KCl	180 mM	6.71 g
EDTA	4 mM	0.745 g
BSA	1 g/l	0.5 g

Adjust pH to 7.4 with Tris, HCl

Isolation buffer B:

Isolation buffer A with 2.5 mg/ml Subtilisin. Add 25 mg Subtilisin to 10 ml of Buffer A.

Isolation buffer C:

Isolation buffer A without BSA

For 3 - 4 isolations 500 ml of media need to be prepared. Add BSA to 250 ml of medium and leave the residual 250 ml without BSA.